

The Formation and Stability of Flavans with 2,3-cis-3,4-cis Configuration

Johan Coetzee,* ,a Elfranco Malan*,a and Daneel Ferreira*,b

^aDepartment of Chemistry, University of the Orange Free State, P.O. Box, 339, Bloemfontein, 9300 South Africa

^bNational Center for Natural Products Research, Research Institute of Pharmaceutical Sciences, School of Pharmacy, The University of Mississippi, University, MS 38677

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Abstract: Treatment of 4β -chloroepioritin tri-O-methylether with phenylmethanethiol under neutral conditions affords both the 4α - and 4β -substituted epioritin derivatives. The stereochemical course of the reaction is of relevance to the thiolysis of 5-oxy (A-ring) proanthocyanidins and to the conspicuous stability of the 7,8-dihydroxy-2,3-cis-3,4-cis-flavan-3,4-diols, teracacidin and melacacidin, and of some of their all-cis (C-ring) oligomers. © 1999 Elsevier Science Ltd. All rights reserved.

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INTRODUCTION

Few reactions, if any, have had a more profound effect on the structure elucidation of the 5-oxy (A-ring) proanthocyanidins than their acid-catalyzed degradation using mercaptans^{1,2} and phloroglucinol³ as capture nucleophiles for the C4-carbocations originating from the chain extender flavan-3-ol moieties. The axial C3-hydroxyl group of extender units with 2,3-cis configuration controls the stereochemical course of coupling of the nucleophile to the transient carbocation leading in all instances but one⁴ to C4-substituted flavan-3-ol units with 3,4-trans configuration.

We recently converted epioritin- 4α -ol tri-O-methyl ether 1 into the 4β -chloroflavan-3-ol derivative 2 and subsequently used the latter compound with its axial C3-OH function as the potential electrophilic flavanyl unit to synthesize a series of novel (4-O-4)- and (4-O-3) bis-teracacinidins^{5,6} via coupling with the C3- or C4-hydroxyl groups of flavan-3,4-diols, e.g. 1. The stereochemical course of the formation of the (4-O-4)-3,4-trans (C-ring) bis-teracacinidin derivative 3 was attributed to an anticipated neighbouring group mechanism.^{6,7} The trans-diaxial arrangement of the C3-hydroxyl group and the C4-chloro nucleofuge of the 4β -chloroflavan-3-ol 2 permits the formation of a transient protonated epoxide 4 which is prone to nucleophilic attack from the sterically less screened β -face. The unexpected formation of the (4-O-4)-3,4-cis (C-ring) bis-teracacinidin 5 was ascribed to conditions incapable of triggering a neighbouring group mechanism hence resulting in an S_N2-type coupling. The generation of the 3,4-cis (C-ring) oxyflavanyl

^{*} Corresponding authors. E-mail: dferreir@olemiss.edu; coetzeej@cem.nw.uovs.ac.za

compound 5,⁶ the formation of the all-*cis* flavan-3,4-diol 1 *via* solvolysis of the 4 β -chloroflavan-3-ol 2 and genesis of 4 α -benzylsulfanylepicatechin 6 in low concentrations during acid-catalyzed thiolysis of procyanidins with epicatechin-(4 β \rightarrow 6)- and (4 β \rightarrow 8)-catechin structural units⁴ prompted investigation of the thiolysis reaction of the 4 β -chloroflavan-3-ol 2 to probe the mechanism of the formation of these all-*cis* flavan derivatives.

RESULTS AND DISCUSSION

Treatment of a solution of the 2,3-cis-3,4-trans-4 β -chloroflavan-3-ol 2⁵ in anhydrous THF with phenylmethanethiol (2.0 eq.) at room temperature (ca 22°C) for 15h, afforded a mixture (68% yield) comprising the 4 α - and 4 β -benzylsulfanylepioritin derivatives 8 (11%) and 9 (89%). These compounds were identified as the 3-O-acetyl derivatives 10 and 11 with ¹H NMR coupling constants indicating 2,3-cis-3,4-cis (J_{2,3} = 1.0; J_{3,4} = 4.0 Hz) and 2,3-cis-3,4-trans (J_{2,3} = 1.5; J_{3,4} = 2.5 Hz) configurations for 10 and 11, respectively. ^{4,8} Phase sensitive NOESY experiments confirmed these configurations via the selective association between 2- and 4-H(C) for the all-cis derivative 10 only. High-amplitude negative ([θ]_{244.6} -1.96x10⁴) and positive ([θ]_{245.8} +1.05x10⁵) Cotton effects in the CD spectra of 10 and 11, respectively, further validated the 4 α - and 4 β -benzylsulfanyl substituents in 10 and 11, resp.

The ca. 1:9 ratio of 3,4-cis- and 3,4-trans-4-benzylsulfanylepioritin derivatives 8 and 9 contrasts with the ca. 1:35 – 1:45 ratio of 4α - and 4β -benzylsulfanylepicatechin analogues 6 and 7 formed in the acid-catalyzed thiolysis of the aforementioned procyanidins.⁴ Since the degradation reactions were performed at the reflux temperature of a mixture of ethanol, phenylmethanethiol and acetic acid, the ratio of 4α - and 4β -substituted products may be temperature dependent. However, treatment of the 4β -chloroflavan-3-ol 2 with phenylmethanethiol in THF for 15h at -60°C and 60°C consistently gave the same ca. 1:9 ratio of 4α - and 4β -benzylsulfanylepioritin derivatives 8 and 9 in similar (ca. 70%) yields.

These results presumably indicate the simultaneous formation of the 4α - and 4β -benzylsulfanylepioritins 8 and 9. The 4α -isomer 8 forms via S_N2 attack of the powerful sulphur nucleophile at the σ^* -orbital of the polarized near axial C4-Cl bond. The proper alignment for this process is attained by intermolecular hydrogen bonding between the axial C3-OH and the thiol as is depicted in 12. A sufficiently polarized C4-Cl bond may trigger the neighbouring group mechanism^{5,6} to permit the formation of the 4β -benzylsulfanylepioritin 9 via intermediacy of the protonated oxirane 4. The relative rate of the anchimeric process must exceed that of the S_N2 route to explain the dominant formation of the 4β -analogue 9.

The thermodynamic stabilities of the 4-thiobenzyl ethers 8 and 9 may also influence the ratio in which they are formed. The less stable 4α -benzylsulfanylepioritin 8 may be protonated at sulphur by the acid generated during thiolysis of the 4β -chloroflavan-3-ol 2. Owing to conformational restraints (see below) the protonated species will then permit a slow conversion into the thermodynamically more stable 4β -thioether 9.

The latter compound should resist C-4 epimerization since enhancement of the nucleofugal properties of the 4-thio functionality by protonation would simply lead to 'activation' of the neighbouring group effect and hence shielding of the α -face at C-4 of oxirane 4. When the 4α -thiobenzyl ether 8 and phenylmethanethiol in dry THF containing gaseous HCl were stirred at 50° C, the all-*cis* compound was indeed slowly converted into the 4β -benzylsulfanylepioritin 9, affording a *ca.* 2:1 mixture of analogues 8 and 9 after 3 days. The 4β -thiobenzyl derivative 9 was stable under the same conditions.

A number of factors, e.g. the strength of the C4-leaving group bond and the nature of the nucleophile, will influence the energetics of the substitution process and hence the competition between the anchimeric and S_N2 processes. This is also evident from the preferential formation of the 'anchimeric product' 7 in the acid-catalyzed thiolysis of selected procyanidins⁴ at ca. 80° C and the selective formation of the all-cis diol 1 during solvolysis of 2. The dual nature of the mechanism leading to the formation of 4α - and 4β -epimers during substitution reactions at C-4 of flavans with 3-axial hydroxyl groups is thus firmly established.

The conspicuous resistance of the 4α -thiobenzyl ether 8 to epimerization at C-4 is related to the observed stability of melacacidin 13 under solvolytic conditions. Such inertia towards solvolysis or epimerization was ascribed^{7,9} to hydrogen bonding between the axial C3-OH and the heterocyclic oxygen which locks the C-ring in a half-chair conformation with C_{2eq} ; C_{3ax} ; C_{4eq} substituents. In this conformation the appropriate C4 σ^* -orbital is at an angle of ca. 45° above the plane of the A-ring and 'buried' in the heterocyclic ring which screens its overlap by an external nucleophile. Since a C-4 antibonding orbital orthogonal to the A-ring would permit the most effective delocalization of A-ring electron density or stabilization of electron deficiency at C-4, it is now clear why an all-cis C-ring configuration is more common for flavonoids with 7,8-dihydroxylated A-rings. These compounds no doubt, would have a reduced need for delocalization $^{10-12}$ of the aromatic electron density than their counterparts with more electron-rich resorcinol-and phloroglucinol-type A-rings. It may then also explain the stability and abundance of the flavan-3,4-diol, teracacidin (free phenolic form of 1) as well as the growing number of dimers with 2,3-cis-3,4-cis-flavanyl constituent units all possessing 7,8-dihydroxy A-rings and axial C-3 hydroxyl groups. $^{5,6,13-16}$

EXPERIMENTAL

¹H NMR spectra were recorded at 298K on a Bruker AM-300 spectrometer for solutions in CDCl₃ with the solvent as internal standard. Mass spectra were recorded on a VG-70-70E spectrometer and CD spectra on a Jasco J-710 spectropolarimeter. Preparative plates (PLC), 20x20 cm, Kieselgel PF₂₅₄ were air dried and used without prior activation. Methylations were performed with an excess of diazomethane in MeOH-Et₂O over 48h at −15°C and acetylations were in acetic anhydride-pyridine at 25°C.

Formation of the 4α - and 4β -benzylsulfanylepioritin derivatives **8** and **9** at 22° C.

A solution of (2R,3S,4S)-2,3-cis-3,4-trans-4-chloro-3-hydroxy-7,8,4'-trimethoxyflavan **2** (95 mg) and phenylmethane thiol (0.068 ml) in dry THF (10 ml) was stirred for 15h at 22°C. The volume was reduced in a

stream of N₂ and the products separated by PLC in benzene-Me₂CO (9:1). The ensuing bands [8, R_f 0.68 (8.7 mg); 9, R_f 0.49 (76.2 mg)] were acetylated and purified by PLC in benzene-Me₂CO (19:1) to give products, respectively, at R_f 0.64 (9.6 mg) and 0.67 (80.4 mg). The R_f 0.64 band comprises (2*R*,3*S*,4*R*)-2,3-*cis*-3,4-*cis*-3-acetoxy-4-benzylsulfanyl-7,8,4'-trimethoxyflavan 10 as a *white amorphous solid*, $\delta_{\rm H}$ 7.47-7.28 (m, ArCH₂, 5xH), 7.40 (d, J = 9.0 Hz, H-2',6'), 7.31 (d, J = 9.0 Hz, H-5), 6.94 (d, J = 9.0 Hz, H-3',5'), 6.57 (d, J = 9.0 Hz, H-6), 5.59 (dd, J = 1.0, 4.0 Hz, H-3), 5.13 (br.s, J = ca 1.0 Hz, H-2), 4.27 (d, J = 4.0 Hz, H-4) 3.92 and 3.87 (each d, J = 13.5 Hz, ArCH₂), 3.89, 3.87, 3.84 (each s, 3xOMe) and 1.99 (s, OAc); m/z, M⁺, 480. (Found: M⁺, 480.1606. C₂₇H₂₈SO₆ requires M, 480.1606); CD [θ]_{278.7} -5.9x10³, [θ]_{263.6} -3.5, [θ]_{244.6} -1.96x10⁴, [θ]_{235.7} +1.35x10³, [θ]_{230.7} -7.85x10²; IR (cm⁻¹, CHCl₃): 1739, 1714, 1610, 1526, 1514.

The R_f 0.67 band afforded (2R,3S,4S)-2,3-cis-3,4-trans-3-acetoxy-4-benzylsulfanyl-7,8,4'-trimethoxyflavan **11** as a white amorphous solid, δ_H 7.51-7.29 (m, ArCH₂, 5xH), 7.36 (d, J = 9.0 Hz, H-2',6'), 6.92 (d, J = 9.0 Hz, H=3',5'), 6.76 (d, J = 9.0 Hz, H-5), 6.54 (d, J = 9.0 Hz, H-6), 5.57 (br.s, J = ca 1.0 Hz, H-2), 5.29 (dd, J = 1.0, 2.5 Hz, H-3), 3.92 and 4.11 (each d, J = 14.0 Hz, ArCH₂), 3.95 (d, J = 2.5 Hz, H-4), 3.89 (s, OMe), 3.85 (s, 2xOMe), 1.90 (s, OAc); m/z, M⁺ 480 (Found: M⁺, 480.1605. C₂₇H₂₈SO₆ requires M⁺, 480.1606); CD [θ]_{290.2} -7.4x10³, [θ]_{262.9} +6.66x10³, [θ]_{245.8} +1.05x10⁵, [θ]_{236.6} -2.83x10³, [θ]_{230.3} +1.09x10⁴, [θ]_{220.6} +8.8x10³; IR (cm⁻¹, CHCl₃): 1734, 1714, 1614, 1540, 1530, 1516.

Similar treatment of the 4β -chloroepioritin derivative 2 (101 mg) for 15h at -60° C afforded 8 (8.9 mg) and 9 (84.3 mg). At 60° C the 4β -chloroepioritin 2 (120 mg) was converted into a mixture comprising 8 (6.4 mg) and 9 (103.2 mg).

Assessment of the stability of the 4α -thiobenzyl ethers 8 and 9.

The thiobenzyl ether **8** (6.3 mg) was dissolved in dry THF (3 ml) containing gaseous HCl [taken from a stock solution of THF which was purged for 30 min with dry HCl (g)]. Phenylmethanethiol (excess) was added, the mixture was stirred at 50° C for 3 days under N_2 and evaporated to dryness. The mixture was resolved by PLC in benzene-Me₂CO (9:1) to give the 4α -benzylsulfanylepioritin derivative **8** (3.2 mg, R_f 0.68) and a further band at R_f 0.49 (2.3 mg). The latter band was acetylated and purified by PLC in benzene-Me₂CO (19:1) to give the 3-O-acetyl-4 β -benzylsulfanylepioritin derivative **11** (1.5 mg, R_f 0.67).

The 4β-benzylsulfanylepioritin derivative 9 was recovered unchanged after similar treatment.

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REFERENCES

1. Betts, M.J.; Brown, B.R.; Brown, P.E.; Pike, W.T. Chem. Comm., 1967, 1110-1112.

- 2. Thompson, R.S.; Jacques, D.; Haslam, E.; Tanner, R.J.N. J. Chem. Soc., Perkin Trans. 1, 1972, 1387-1399
- 3. Foo, L.Y.; Porter, L.J. J. Chem. Soc., Perkin Trans. 1, 1978, 1186-1190.
- 4. Kolodziej, H. Phytochemistry, 1990, 29, 1671-1674.
- 5. Coetzee, J.; Malan, E.; Ferreira, D. J. Chem. Res., 1998, (S) 526-527; (M) 2287-2296.
- 6. Coetzee, J.; Malan, E.; Ferreira, D. Tetrahedron, 1998, 54, 9153-9160.
- 7. Clark-Lewis, J.W.; Williams, L.R. Aust. J. Chem., 1967, 20, 2152-2167.
- 8. Clark-Lewis, J.W. Aust. J. Chem., 1968, 21, 2059-2075.
- 9. Clark-Lewis, J.W.; Mortimer, P.I. J. Chem. Soc., 1960, 4106-4111.
- 10. Malan, E.; Roux, D.G. Phytochemistry, 1975, 1835-1841.
- 11. Roux, D.G.; Ferreira, D. in 'Progress in the Chemistry of Natural Products', eds. Herz, W.; Grisebach, H.; Kirby, G.W., Springer-Verlag, Berlin, 1982, 41, 47-76.
- 12. Foo, L.Y. J. Chem. Soc., Chem. Commun., 1985, 1273-1274.
- 13. Foo, L.Y. J. Chem. Soc., Chem. Commun., 1986, 236-237.
- 14. Foo, L.Y. J. Chem. Soc., Chem. Commun., 1989, 1505-1506.
- 15. Malan, E. Phytochemistry, 1995, 38, 237-239.
- 16. Malan, E.; Sireeparsad, A.; Swinny, E.; Ferreira, D. Phytochemistry, 1997, 44, 529-531.